

Associations of MYPN, TTN, SCN5A, MYO6 and ELN Mutations With Arrhythmias and Subsequent Sudden Cardiac Death: A Case Report of an Ecuadorian Individual

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Abstract

Cardiac pathologies are among the most frequent causes of death worldwide. Regarding cardiovascular deaths, it is estimated that 5 million cases are caused by sudden cardiac death (SCD) annually. The primary cause of SCD is ventricular arrhythmias. Genomic studies have provided pathogenic, likely pathogenic, and variants of uncertain significance that may predispose individuals to cardiac causes of sudden death. In this study, we describe the case of a 43-year-old individual who experienced an episode of aborted SCD. An implantable cardioverter defibrillator was placed to prevent further SCD episodes. The diagnosis was ventricular fibrillation. Genomic analysis revealed some variants in the MYPN (pathogenic), GCKR (likely pathogenic), TTN (variant of uncertain significance), SCN5A (variant of uncertain significance), MYO6 (variant of uncertain significance), and ELN (variant of uncertain significance) genes, which could be associated with SCD episodes. In addition, a protein-protein interaction network was obtained, with proteins related to ventricular arrhythmia and the biological processes involved. Therefore, this study identified genetic variants that may be associated with and trigger

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Keywords: Ventricular arrhythmias; Sudden death; Mutations; NGS

Introduction

Sudden cardiac death (SCD) refers to the unexpected death of an individual not attributable to an extracardiac cause. SCD usually happens within the first hour from the onset of symptoms or within 24 h from the last time the individual was seen in good health [1, 2]. It is estimated that approximately 5 million cases of SCD occur worldwide each year [3]. Furthermore, the survival rate is between 27% and 40%, and this percentage has increased due to factors such as cardiopulmonary resuscitation capabilities and external defibrillators. Various causes could trigger SCD, including coronary artery anomalies, structural cardiac anomalies, primary arrhythmias, and coronary artery disease, among others [4].

Cardiac arrhythmias lead to more than one million cases of syncope yearly and are associated with congenital, metabolic, structural, physiological, immunological, and infectious disorders [5, 6]. Among these arrhythmias, ventricular fibrillation (VF) is responsible for the highest number of cases of sudden cardiac arrest (SCA) [7]. VF is a disturbance in the electrical activity of the heart's lower chambers that results in the absence of an effective heartbeat [8].

Diagnosing arrhythmogenic abnormalities could be complex, as they may manifest as minimal findings in a morphologically normal heart [9]. Moreover, the identification of substrates and triggers of arrhythmias has provided insights into the molecular basis of different pathophysiological pathways [10]. Arrhythmias are diseases with genetic heterogeneity, and as a result, genomic analysis has played a crucial role in the diagnosis of primary arrhythmic disorders by uncovering muta-

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Figure 1. Subject's episodes of care. The relevant data are displayed in the timeline.

tions in different genes correlated with the same disorder [11].

This study describes the case of a man who suffered an aborted SCD, and through genomic analysis, our research group identified variants in genes associated with this cardiac condition.

Case Report

Investigations

A 43-year-old Ecuadorian healthy man, with no history of heart disease, suffered an SCD at his workplace 7 years ago. Emergency services resuscitated him, and he was hospitalized. In the hospital, electrocardiogram and echocardiogram were performed, but no structural cardiac cause was found to be associated with the SCD event. The cardiac condition diagnosed was primary VF. Unfortunately, the patient did not respond to medications satisfactorily; thus, an implantable cardioverter defibrillator (ICD) was placed to prevent another sudden death episode. The implanted ICD detected several sudden death events; however, the evolution of the individual's cardiac condition was favorable. Furthermore, the patient was prescribed an antiarrhythmic drug, amiodarone 200 mg, as a regular medication.

Five years after the SCD incident, the individual was hospitalized due to new syncope events, palpitations, precordial pain, and dyspnea in functional class II/IV. Subsequently, electrocardiogram and echocardiogram were performed, revealing the need for an elective ICD replacement. The discharge diagnoses included New York Heart Association (NYHA) class III heart failure, hemodynamic profile B, and congestive heart failure stage C due to tachycardiomyopathy. A timeline of the relevant episodes of care is depicted in Figure 1.

Diagnosis

Next-generation sequencing (NGS)

A peripheral blood sample was taken, and DNA was extracted using the PureLink[™] genomic DNA mini kit. DNA concentrations were quantified using the 1X dsDNA high-sensitivity (HS) and broad range (BR) assay kits on the Qubit[™] 4 fluorometer. NGS was performed at the Centro de Investigacion Genetica y Genomica (CIGG) using the TruSight[™] cardio (TSC) sequencing panel on the Illumina MiSeq platform. The TSC sequencing panel includes 174 genes with known associations with 17 inherited cardiovascular conditions. For the bioinformatics analyses, DRAGEN Enrichment v3.9.5, Annotation Engine v3.15, PolyPhen, Sift, and Variant Interpreter v2.16.1.300 platforms were used.

Ancestral components determination

Forty-six ancestry-informative INDEL markers (AIMs) were amplified in a multiplex PCR reaction, according to Zambrano et al (2019). Fragment detection was performed on the 3500 genetic analyzer. The results were collected and analyzed on the Data Collection v3.3 and Gene Mapper v.5 platforms. The ancestral analysis was performed using STRUCTURE v.2.3.4 [12].

Outcomes

The coverage was \geq 50 X on 98.19% of the target regions of the TSC sequencing panel. Variants were classified into five categories (benign, likely benign, variants of uncertain significance (VUS), likely pathogenic, and pathogenic) following the 2015 American College of Medical Genetics and Genomics -Association for Molecular Pathology guidelines [13]. All pathogenic, likely pathogenic, and VUS variants were considered in the analysis (Table 1).

To investigate the cellular processes involving the mutated proteins, protein-protein interaction networks were created using the STRING database [14], with a confidence level of 0.7 $(P < 1.0 \times 10^{-16})$. The network was generated using 34 genes related to the individual's phenotype and found in the arrhythmia sequencing panel [15] (Supplementary Material 1, www. cardiologyres.org). Notably, the TSC sequencing panel and the arrhythmia sequencing panel share 24 genes. Subsequently, it was determined that of the six genes that presented mutation, five were related to arrhythmias. Among these five genes, only the *TTN* and *SCN5A* were present in both sequencing panels.

Furthermore, it has been established that the MYPN protein physically interacts with the TTN protein. Moreover, both the TTN and the SCN5A proteins interact with the CALM1 protein. All these proteins have a physical interaction with other arrhythmia-causing proteins and are involved in the actin filament-based processes (Fig. 2).

Moreover, an ancestral composition analysis was performed, and the results showed 2% African, 32.8% European, and 65.2% Native American components.

Discussion

In this case report, genomic analyses of a 43-year-old Ecuadorian male with primary VF and subsequent episodes of aborted sudden death were performed. The objective was to determine the association between genetic variants and the individual's phenotype. Our study identified five mutated genes related to arrhythmias, as well as an unrelated gene. The genomic screening revealed variants in the MYPN, TTN, SCN5A, MYO6, and ELN genes. These variants could show an association between genetic factors, arrhythmias disorders and the increased risk of SCD.

MYPN

Mutations in the sarcomeric myopalladin (MYPN) protein play a significant role in the pathogenesis of cardiac disease. MYPN interacts with several molecules, including α -actinin, located along the stress fibers and in the Z-line of cardiac muscles. Furthermore, the MYPN protein functions as a cytoskeleton

lable 1	و -	lable 1. Genetic variants identified Using I SC Sequencing Panel	sequencing Panel				
Gene		Chr HGVSP DNA reference	HGVS protein reference	Consequence	Predicted effect	Predicted effect dbSNP/dbVar ID Genotype	Genotype
MYPN	10	MYPN 10 NM_001256267.1; c.3335C>T	NM_001256267.1; p.(Pro1112Leu)	Missense variant	Pathogenic	rs71534278	Heterozygous
GCKR	2	NM_001486.3; c.307G>A	NM_001486.3; p.(Val103Met)	Missense variant	Likely Pathogenic rs146175795	rs146175795	Heterozygous
TTN	7	NM_001267550.2; c.71641G>C	NM_001267550.2; p.(Val23881Leu)	Missense variant	NUS	N/A	Heterozygous
TTN	2	NM_001267550.2; c.9670A>G	NM_001267550.2; p.(Arg3224Gly)	Missense variant	NUS	rs557221911	Heterozygous
SCN5A 3	б	NM_198056.2; c.2302A>G	NM_198056.2; p.(lle768Val)	Missense variant	NUS	N/A	Heterozygous
90AW	9	NM_004999.3; c.2506C>T	NM_004999.3; p.(Arg836Cys)	Missense variant	NUS	rs138305791	Heterozygous
ELN	Г	NM_001278939.1; c.2142_2156del	NM_001278939.1; c.2142_2156del NM_001278939.1; p.(Gly715_Val719del) In-frame deletion variant	In-frame deletion variant	NUS	rs782015142	Heterozygous
110.10	10	// I.C					

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/US: variants of uncertain significance



Figure 2. Protein-protein interaction network between the arrhythmia panel genes and the mutated genes on the TSC sequencing panel. The purple circle nodes represent the actin filament-based process. The yellow circle nodes represent cell communication involved in cardiac conduction. The solid lines indicate the physical interaction between proteins, whereas the red circle shows the genes analyzed in the genomic screening. TSC: TruSight[™] cardio.

support and signaling mediator [16]. Mutations in this protein can affect both contractile (myosin, actin) and non-contractile (titin, Z-disc proteins) proteins of the sarcomere [17].

According to the literature, mutations in the MYPN protein are associated with different types of cardiomyopathies, such as hypertrophic, dilated, and restrictive. In addition, some studies suggest that the *MYPN* gene network is involved in specific arrhythmia disorders or overlapping phenotypes of inherited cardiomyopathies [18, 19]. However, these types of mutations are rare, accounting for less than 5% [20]. In this genomic screening, a p.(Pro1112Leu) missense MYPN pathogenic variant was described. Although the replacement of the amino acid proline by leucine in this mutation may not be pathogenic, it is essential to consider that the coexistence of other mutations could impact the phenotype [16]. Importantly, the Ecuadorian mestizo individual in this study has not been diagnosed with any of the previously mentioned cardiomyopathies.

SCN5A

The NGS results showed a mutation of uncertain significance at position c.2302A>G (p.Ile768Val) in exon 15 of the *SCN5A* gene. The *SCN5A* gene encodes the sodium channel-forming alpha subunit that regulates sodium influx and is involved in the rapid upward depolarization of the action potential. The SCN5A protein, with a molecular weight of 227 kDa, consists of four homologous domains (DI-DIV), and each one is composed of six segments (S1-S6) [21]. Mutations in *SCN5A* have been associated with various cardiac diseases, such as ventricular arrhythmias, sudden death syndrome, and cardiac conduction disturbances [22].

ELN

The human elastin gene, *ELN*, is part of the extracellular matrix and is involved in the elasticity and strength of tissues such as arterial vessels, lungs, and others [23]. Elasticity allows blood vessels to perform the process of diastole and systole for optimal heart function. In addition, in arteries, elasticity maintains tone and regulates blood pressure [24].

Mutations in the *ELN* gene are associated with diseases such as supravalvar aortic stenosis (SVAS), characterized by significant narrowing of the large arteries [25, 26]. In the genomic analysis performed in the individual, a mutation of uncertain significance was identified at position c.2142_2156 of (p.Gly715_Val719del), resulting in an in-frame deletion in exon 30 of the *ELN* gene. Mutations in the *ELN* gene may be associated with a sporadic or inherited autosomal dominant SVAS.

In 2016, Latham et al mentioned that the SCD risk in patients with SVAS is increased compared to non-syndromic SVAS patients [27]. Therefore, it is essential to understand this type of mutation in patients diagnosed with SVAS, particularly

for medical examinations or surgeries that require sedation, since there have been cases of SCD leading to the fatal outcomes [23].

MYO6

MYO6 gene encodes a protein that moves toward the minus end of actin filaments. The protein has a motor domain with ATP and actin-binding sites and a globular tail that interacts with other proteins. Mutations in the motor domain of myosin could cause cardiac problems, such as dilated and hypertrophic cardiomyopathies [28], as well as different forms of congenital heart defects [29]. However, our patient does not present any of these alterations.

TTN

The *TTN* gene encodes a large and abundant protein found in striated muscle. This protein is divided into two regions: the N-terminal I-band, which is the elastic part, and the C-terminal A-band that acts as a regulatory protein. The N-terminal region of the Z-disk and the C-terminal region of the M-line bind to the Z-line and the M-line of the sarcomere, respectively. Additionally, TTN contains binding sites for muscle-associated proteins, contributing to the assembly of the contractile machinery in muscle cells [30].

TTN variants are the leading cause of dilated and familial hypertrophic cardiomyopathies and have also been associated with an increased risk of ventricular arrhythmias [31]. However, it is important to highlight that our patient does not present any of these cardiomyopathies.

GCKR

The heterozygous mutation p.(Val103Met) of the glucokinase regulatory gene (*GCKR*) is another variant found in this study as likely pathogenic. The *GCKR* gene encodes the glucokinase regulatory protein (GCK), which acts as a glucose regulator in hepatocytes and β cells of the pancreas [32, 33].

According to a kinetic assay characterization of the p.Val103Met variant, it was classified as a severe loss-of-function variant due to its ability to inhibit GCK protein activity [32]. This mutation, p.Val103Met, was found in non-Hispanic individuals of mixed European ancestry, according to the study by Rees et al (2012). While some research describes the association of GCK function with coronary artery disease and ischemic stroke [34], there is no evidence of an association with arrhythmias in the literature.

Protein-protein interaction network

The protein-protein interaction network of physical protein interactions and biological processes can provide valuable information about the relationship with arrhythmia processes. For instance, the actin filament-based biological process, as shown in Figure 2, has been described by Camors et al (2022). The authors mention that changes in actin protein expression are related to the progressive reduction of the right ventricular contraction and development of arrhythmias in a premature phase of arrhythmogenic cardiomyopathy [35].

In some genetic cardiomyopathies, clinical variability is observed even among patients with similar genotypes, ranging from asymptomatic cases to heart failure. This heterogeneity suggests that other factors play a significant role in modifying the clinical phenotype, potentially influencing disease potentiation or protection. These factors include modulatory genes, polymorphism, other unknown genes, as well as environmental and endogenous factors (age, sex, physical exercise, drugs, hormones, viral infections, and emotional stress) [36].

Furthermore, studies suggest that the combination of genetic variants and the additive effect of mutations in different genes could potentially trigger a more aggressive phenotype in the context of cardiac arrhythmias [37].

Conclusion

SCD is often caused by cardiac disorders that can remain asymptomatic and difficult to diagnose. Genomic screening is a valuable tool that could reveal pathogenic, likely pathogenic, and VUS genetic variants related to a phenotype. The genetic variants identified in the Ecuadorian individual, particularly in the *MYPN*, *GCKR*, *TTN*, *SCN5A*, *MYO6*, and *ELN* genes, may be related to cardiac diseases that could be associated with primary VF and SCD. It is also possible that the genetic variants found could have an additive effect on the development of arrhythmia diseases.

Therefore, the identification of these variants could significantly contribute to the comprehension of the genetic basis of arrhythmias and, by extension, SCD. In addition, it would be important to expand the study to encompass other individuals afflicted with the same cardiac condition, to ascertain any potential association with the genetic variants examined in this research article.

Supplementary Material

Suppl 1. Genes and phenotypes associated of the arrhythmia and TruSight[™] Cardio sequencing panels.

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Conflict of Interest

The authors declare no conflict of interest.

Informed Consent

The participant provided their written informed consent to participate in this study. The study conducted with human participants followed the ethical standards of the 2013 Helsinki Declaration and was approved by the Committee on Ethics and Research in Human Subjects (CEISH)-UTE University (protocol code CEISH-2021-016, date of approval 18-05-2022).

Author Contributions

Conceptualization: EPC, VARP and AKZ. Methodology: EPC, VARP, AKZ, SCU, PGR, RTT, JLLB, ND, AAIR, and POR. Writing - original draft preparation: EPC and VARP. Writing - review and editing: EPC, VARP and SCU. Supervision: AKZ. Project administration: AKZ. Funding acquisition: AKZ. All authors read and approved the final manuscript.

Data Availability

The data supporting the findings of this study are available from the corresponding author upon reasonable request.

Abbreviations

AIMs: ancestry-informative INDEL markers; GCKR: glucokinase regulatory gene; HS: high-sensitivity; ICD: implantable cardioverter defibrillator; MYPN: sarcomeric myopalladin; NGS: next-generation sequencing; NYHA: New York Heart Association; SCA: sudden cardiac arrest; SCD: sudden cardiac death; SVAS: supravalvar aortic stenosis; TSC: TruSightTM Cardio; VF: ventricular fibrillation; VUS: variant of uncertain significance; ACS: acute coronary syndrome; BMI: body mass index; CABG: coronary artery bypass grafting; CAD: coronary artery disease; CKD: chronic kidney disease; COPD: chronic obstructive pulmonary disease; ESRD: end-stage renal disease; HIV: human immunodeficiency virus; LAD: left anterior descending; PCI: percutaneous coronary intervention; PE: pulmonary embolism; STEMI: ST-elevation myocardial infarction

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